

# Source Attribution of Poly- and Perfluoroalkyl Substances (PFASs) in Surface Waters from Rhode Island and the New York Metropolitan Area

Xianming Zhang<sup>\*,†,‡</sup>, Rainer Lohmann,<sup>§</sup> Clifton Dassuncao,<sup>†,‡</sup> Xindi C. Hu,<sup>†,‡</sup> Andrea K. Weber,<sup>†</sup> Chad D. Vecitis,<sup>†</sup> and Elsie M. Sunderland<sup>†,‡</sup>

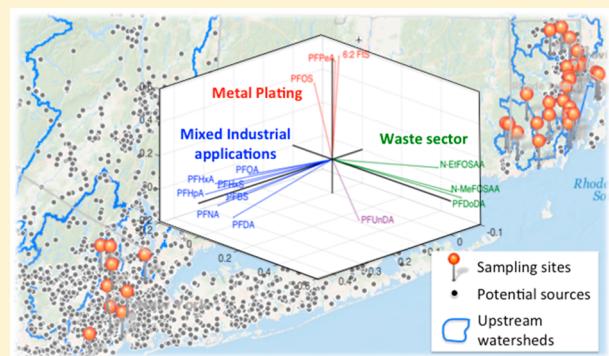
<sup>†</sup>Harvard John A. Paulson School of Engineering and Applied Sciences, Harvard University, Cambridge, Massachusetts 02138, United States

<sup>‡</sup>Department of Environmental Health, Harvard T. H. Chan School of Public Health, Harvard University, Boston, Massachusetts 02115, United States

<sup>§</sup>Graduate School of Oceanography, University of Rhode Island, Narragansett, Rhode Island 02882, United States

## S Supporting Information

**ABSTRACT:** Exposure to poly- and perfluoroalkyl substances (PFASs) has been associated with adverse health effects in humans and wildlife. Understanding pollution sources is essential for environmental regulation, but source attribution for PFASs has been confounded by limited information about industrial releases and rapid changes in chemical production. Here we use principal component analysis (PCA), hierarchical clustering, and geospatial analysis to understand source contributions to 14 PFASs measured across 37 sites in the northeastern United States in 2014. PFASs are significantly elevated in urban areas compared to rural sites except for perfluorobutanesulfonate, N-methyl perfluorooctanesulfonamidoacetic acid, perfluoroundecanate, and perfluorododecanate. The highest PFAS concentrations across sites were those of perfluorooctanate (PFOA, 56 ng L<sup>-1</sup>) and perfluorohexanesulfonate (PFHxS, 43 ng L<sup>-1</sup>), and perfluorooctanesulfonate (PFOS) levels are lower than earlier measurements of U.S. surface waters. PCA and cluster analysis indicate three main statistical groupings of PFASs. Geospatial analysis of watersheds reveals the first component/cluster originates from a mixture of contemporary point sources such as airports and textile mills. Atmospheric sources from the waste sector are consistent with the second component, and the metal smelting industry plausibly explains the third component. We find this source-attribution technique is effective for better understanding PFAS sources in urban areas.



## INTRODUCTION

Exposure to poly- and perfluoroalkyl substances (PFASs) has been associated with many negative health outcomes, including compromised immune function, metabolic disruption, obesity, and altered liver function.<sup>1</sup> PFASs in surface waters are an emerging concern for U.S. public water supplies, and long-chain compounds bioaccumulate in aquatic food webs, posing health risks to seafood consumers.<sup>2–6</sup> Production of PFASs and their precursors has shifted dramatically over the past two decades toward shorter-chain and polyfluorinated species.<sup>7</sup> Diverse point sources and atmospheric deposition of some PFASs confound our understanding of the dominant contributors to contamination in the aquatic environment. Regulatory databases such as the U.S. Environmental Protection Agency's Facility Registry Survey (FRS)<sup>8</sup> and the Toxic Release Inventory<sup>9</sup> presently contain limited to no information about the amounts of PFASs released to the environment.

Multivariate statistical analyses based on chemical composition profiles can be a powerful tool for diagnosing

contamination sources, as illustrated for many other organic contaminants.<sup>10</sup> Principal component analysis (PCA) provides information about interrelationships among various chemicals and is useful for deriving common source profiles. Two-way hierarchical clustering can be used as a confirmatory analysis of PCA by generating a flexible number of subgroups of similar sites (those affected by a common source type) without dictating the number of clusters *a priori*. Clustering of compounds identifies chemicals that co-occur to form a unique signature. These techniques have not been routinely applied to interpret PFAS contamination and show potential for interpreting sources in surface water and seawater.<sup>4,11</sup>

Here we combine PCA and hierarchical clustering of PFAS profiles measured in surface waters from 37 rivers, streams and

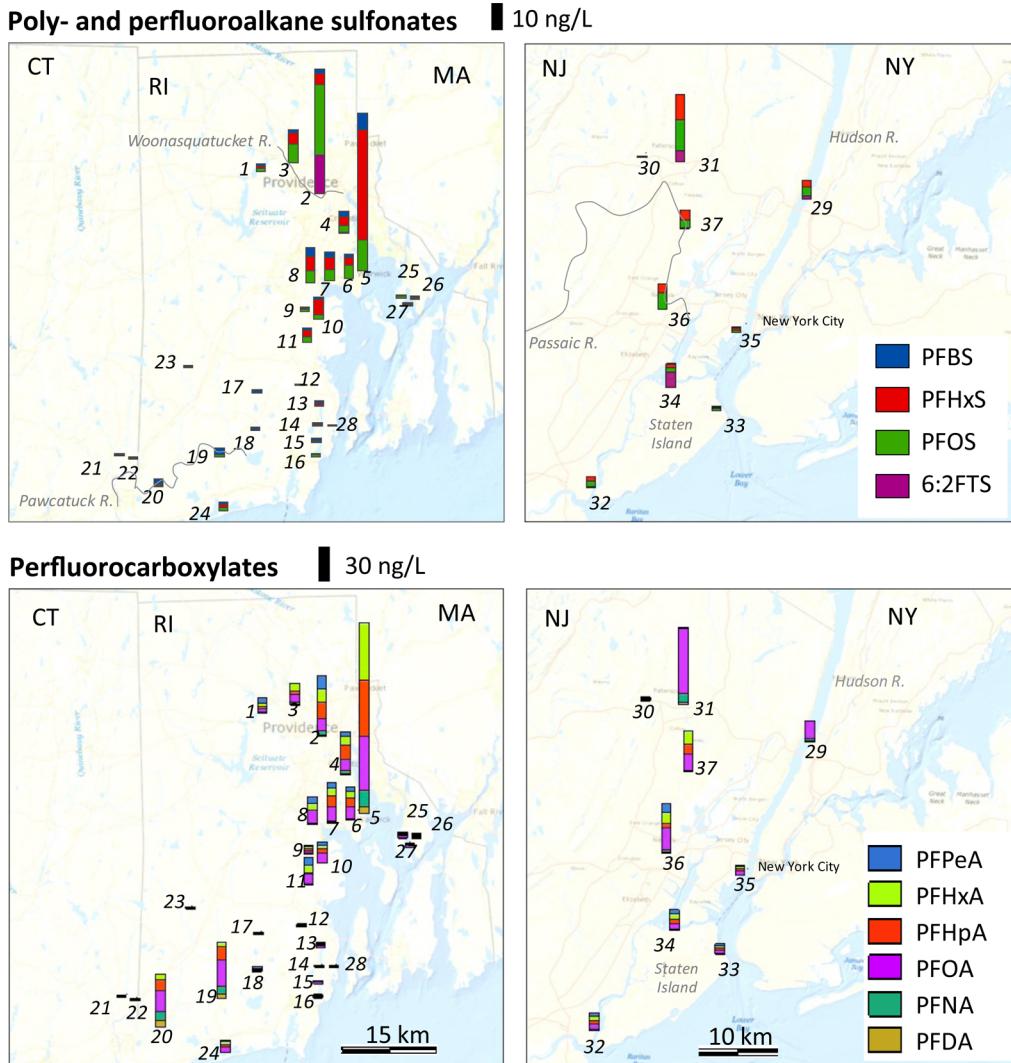
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**Figure 1.** Concentrations of PFASs measured in surface waters from Rhode Island and the New York Metropolitan Area. Full names of individual compounds are listed in [Table S2](#). N-MeFOSAA and N-EtFOSAA are not shown but were detected in ~70% of the samples at concentrations of <1 ng/L.

estuaries in the northeastern United States with geospatial analysis of potential sources. Few measurements of PFASs in U.S. surface waters over the past five years are available, and the importance of different sources is poorly understood. Source regions for air pollution are commonly identified using back trajectories.<sup>12,13</sup> We apply an analogous approach to identify sources of aquatic pollution based on hydrological distances within a watershed. The main objective of this study is to identify major sources of surface water PFAS contamination in diverse watersheds using information about chemical composition and geospatial analytical tools that consider surface hydrology.

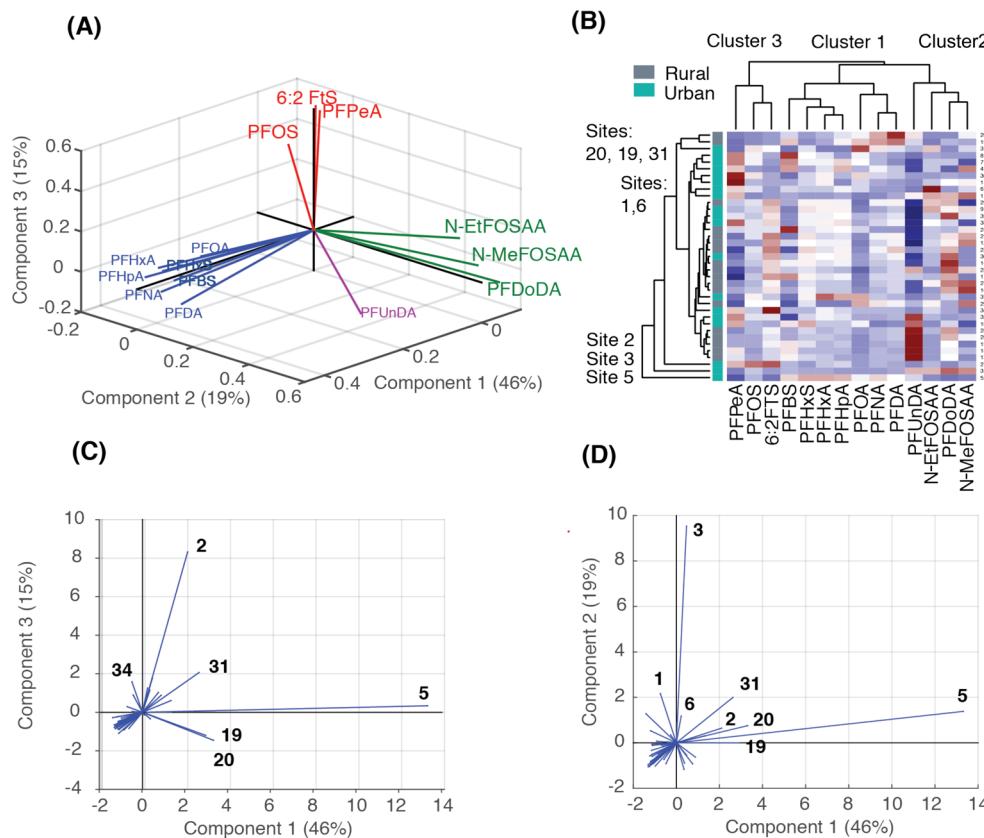
## METHODS

**Sample Collection and Analysis.** We collected surface water samples from rivers, creeks, and estuaries at a depth of approximately 1 m at 28 sites in the state of Rhode Island (RI) in June 2014 and nine sites in the New York Metropolitan Area (NY/NJ) in October 2014 ([Figure 1](#)). A complete description of sampling sites is provided in [Table S1](#). Precipitation and flow rates in rivers tend to be higher in June, potentially resulting in

enhanced dilution and a low bias for some PFASs measured in RI rivers compared to NY/NJ rivers.

Samples were stored in 1 L pre rinsed polypropylene bottles at -20 °C and thawed to room temperature. Each sample was homogenized by shaking vigorously before subsampling 500 ml for the analysis of 21 PFASs. Each unfiltered sample was spiked with 20 µL of a 0.1 ng µL<sup>-1</sup> mass-labeled PFAS mixture (Wellington, Guelph, ON; individual compounds are listed in [Table S2](#)) as internal standards for quantification. PFASs were extracted using an Oasis Wax solid phase extraction (SPE) cartridge (6 mL, 150 mg of sorbent) following the method of Taniyasu et al.<sup>14</sup> (see [Section S1](#) for details). A nitrogen evaporator (ZIPVAP) was used to concentrate the extract to 1 mL [1:1 (v:v) methanol:water].

Sample detection for 21 native PFASs ([Tables S2 and S3](#)) was performed using an Agilent 6460 LC-MS/MS instrument equipped with an online-SPE system (Agilent 1290 Infinity Flex Cube) in dynamic multiple-reaction mode (sample chromatogram in [Figure S1](#)). At least one negative control (field or procedural blank) and one positive control (spiked with 2 ng of the 21 PFASs in 500 mL of water) were included in every extraction batch. Whole method recovery tested using the



**Figure 2.** Multivariate statistical analysis of surface water data. (A) Loadings of principal component analysis (PCA). (C and D) Score plots for three components across sampling sites. (B) Comparison of PCA results to hierarchical clustering of compounds and sites. Sites with statistically distinct PFAS profiles are indicated in panels C and D and highlighted on the hierarchical clustering diagram. The three principal components together explain 80% of the variance in PFAS composition.

positive controls was 70–120% for all but four PFASs that ranged from 60 to 70%, which is comparable to recoveries reported by previous studies.<sup>3,14,15</sup> The four PFASs are perfluoropentanate (PFPeA), perfluoroheptanate (PFHpA), N-methyl perfluorooctanesulfonamidoacetic acid (N-MeFOSAA), and N-ethyl perfluorooctanesulfonamidoacetic acid (N-EtFOSAA). Potential analyte loss during sample preparation was corrected using internal standards spiked prior to sample extraction. The limit of detection [LOD (Figure S2)] was defined as equivalent to the blank plus the concentration corresponding to a signal-to-noise ratio of three. Variability between duplicates obtained at two sites was <20%. Concentrations of PFASs in five field blanks (HPLC grade water) prepared following the sample preparation procedure were all below the LOD.

We quantified branched isomers for perfluorooctanate (PFOA), perfluorohexanesulfonate (PFHxS), perfluorooctane-sulfonate (PFOS), N-MeFOSAA, and N-EtFOSAA using calibration standards for the linear isomers, assuming the same instrumental response factor (Table S3). Seven compounds, namely, perfluorododecanesulfonate (PFDS), 8:2 fluorotelomer sulfonate (8:2 FTS), perfluorooctane sulfonamide (FOSA), and perfluorocarboxylates with more than 12 carbon atoms, were detected in less than half of the samples and were excluded from additional statistical analysis (see Table S2 for details). For the 14 PFASs that had detection frequencies of >60% (Table S2), we used the Robust Regression on Order Statistics approach for censored log-normally distributed

environmental data described by Helsel<sup>16</sup> to assign values to samples with concentrations below the LOD.

**Statistical and Spatial Analysis.** We used PCA and hierarchical clustering to group sites with statistically distinct PFAS composition profiles. PCA was performed using MATLAB's Statistics Toolbox (MathWorks, Inc.) on normalized (z-score to remove the effect of the concentration difference at different sites) PFAS concentration data. The inverses of variances of the data were used as variable weights, and varimax rotation was applied to interpret the meaning of extracted principal components. Hierarchical cluster analysis was conducted using the hclust function in the R statistical computing package (version 3.1.3).

We characterized the watershed for each freshwater sampling site using the U.S. Geological Survey's (USGS) National Elevation Data set (3 arc-second for sites 15 and 16 and 1 arc-second for others) and the Hydrologic Tool in ArcGIS Pro 1.2 and ArcGIS online. Estuarine sampling sites were excluded from the geospatial analysis because of the confounding influence of tidal waters on potential source profiles. Population within each watershed was based on ESRI's U.S. Demographic Database.<sup>17</sup> We used the USGS's StreamStats database (version 4)<sup>18</sup> to characterize water flow rates for each location and to compute mass flow (kilograms per year) of PFASs at each site and per-capita mass flows (kilograms per person per year).

For all inland sites (non-estuarine), we acquired a list and geospatial data for plausible PFAS sources from the FRS database on facilities and sites subject to environmental regulation (see the Supporting Information for the search

criteria).<sup>8</sup> These include airports, facilities for metal plating/coating, printing, sewage treatment, and waste management (including landfills), and manufacturers of semiconductors, textiles, paints/coatings/adhesives, ink, paper, and petroleum products. A caveat of this analysis is that not all facilities included in the FRS database necessarily release PFASs and the database may not comprehensively include all possible sources.

Hydrological distances of point sources from each sampling site were computed using the ArcGIS Trace Downstream tool. Within each watershed, we defined an indicator for the impact of potential point sources as a function of distance from the sampling location by assuming an exponential decay in the source signature<sup>19</sup> (i.e., impact =  $1/e^d$ , where  $d$  is the hydrological distance in kilometers). This approach provides additional information about plausible sources that complements multivariate statistical analysis but cannot be considered a quantitative estimate of contributions to sampling locations because magnitudes of PFAS discharges are not available.

## RESULTS AND DISCUSSION

**Concentrations and Spatial Patterns.** Figure 1 shows the compound specific composition and concentrations of PFASs measured in surface water samples as part of this work. Sampling sites in NY/NJ had population density in upstream watersheds (10 to 43-fold) much greater than the population density of those in RI, but the highest concentrations of most PFASs were measured near the city of Providence, RI (Figure 1 and Figure S2). The range of measured PFAS concentrations reported here is similar to or lower than those of U.S. surface waters from other regions collected between 2000 and 2009 (Table S4).<sup>2,20–24</sup>

All sites had detectable PFOA and PFNA, and >90% contained detectable PFHxS, PFOS, PFDA, and 6:2 FtS (Tables S2 and S3 and Figure S2). The highest individual PFAS concentration across sites was that of PFOA (56 ng L<sup>-1</sup>) at site 31 (Passaic River, NJ). The highest concentrations of PFHxS (43 ng L<sup>-1</sup>) and PFNA (14 ng L<sup>-1</sup>) were measured at site 5 (Mill Cove, RI). The maximal PFOS concentration (27 ng L<sup>-1</sup>) was measured at site 2 (Woonasquatucket River, RI) within the City of Providence, RI. This is much lower than maximal levels reported in earlier studies of U.S. surface waters that ranged between 43 and 244 ng L<sup>-1</sup> (Table S4) and reflects the continued decline in environmental PFOS burdens in North America following elimination of production in 2002.<sup>25,26</sup>

Measured PFAS concentrations in urban regions were significantly higher (Wilcoxon rank sum test;  $p < 0.017$ ) than those at rural sites for all compounds except PFBS, N-MeFOSAA, PFUnDA, and PFDoDA (Figure S3). Sites 1–11 in RI and Sites 29–37 in NY/NJ are all urban areas, defined by population densities of >1000 individuals per square mile (2590 km<sup>2</sup>) and population densities of >500 individuals per square mile in surrounding census blocks.<sup>27</sup> We did not find a statistically significant correlation between the total population in each upstream watershed and PFAS concentrations measured at each sampling site ( $p = 0.12$ –0.95 across compounds). We derived per-capita discharges (Figure S4) using an approach similar to that of Pistocchi and Loos.<sup>28</sup> Highest median per-capita discharges (micrograms per person per day) across compounds, in decreasing order, were for PFOA (27), PFHxA (14), PFHpA (10), PFOS (9), PFHxS (7), and PFNA (5) (Figure S4). These are lower than those previously reported in Europe ca. 2007 (e.g., PFOA, 82 µg person<sup>-1</sup> day<sup>-1</sup>; PFOS, 57 µg person<sup>-1</sup> day<sup>-1</sup>).<sup>28</sup>

**Source Identification.** Both hierarchical clustering and PCA identified three distinct groupings of PFASs (Figure 2A,B). The first component/cluster explains 46% of variability in the PCA and includes two major end products of the fluorochemical manufacturing industry (PFOA and PFNA), and a mix of other compounds (PFBS, PFHxS, PFHxA, and PFDA). Site 5 (Mill Cove, RI) contains the highest summed concentration of PFASs across all sites and is dominated by this mixture of PFASs. PCA results suggest site 5 is statistically similar to the Pawcatuck River, RI, sampling locations (sites 19 and 20) and the Passaic River, NJ, location (site 31). However, these sites are grouped separately in the hierarchical clustering analysis (Figure 2B), suggesting some differences in source contributions.

Geospatial analysis of the watersheds for sites 5, 19, 20, and 31 reveals a mixture of potential sources (Figure S5). For site 5, the greatest source impact as a function of distance within the watershed is from T. F. Green Airport, the largest public airport in Rhode Island. Prior work indicates uses of AFFF at modern airports release diverse PFASs to downstream aquatic environments, including the compounds identified as part of the first PCA/cluster.<sup>4,29–31</sup> For Sites 19 and 20, textile mills in the upstream watersheds have the greatest impact as a function of distance (Table S5). PFASs are used for water resistant coating in textiles, and washing and disposal of wastewater at textile mills provides a vector for their entry to the aquatic environment. For site 31, PCA scores suggest a mix of components 1–3 (Figure 2C,D). This site also clusters differently than sites 19 and 20 (Figure 2B). The FRS database indicates the watershed of site 31 (Figure S5) contains diverse industrial sources that must account for this profile, including metal plating, printing, a landfill, petroleum, and coal product manufacturing. Overall, we conclude that the first PCA component and cluster of PFASs (PFOA, PFNA, PFBS, PFHxS, PFHxA, and PFDA) represent a mixture of contemporary sources, including airports and textile mills.

The second component/cluster explains 19% of the variability in PFASs and includes two long-chain PFASs (PFUnDA and PFDoDA) and two precursors to PFOS (N-MeFOSAA and N-EtFOSAA) (Figure 2). PFUnDA and PFDoDA mainly originate from fluorotelomer alcohols or other fluorotelomer-based products.<sup>32</sup> Both N-MeFOSAA and N-EtFOSAA are intermediate degradation products from the volatile parent compound *N*-alkyl perfluorooctansulfonamidoethanol (FOSE) with PFOS as the final degradation product. This profile is most pronounced at site 3 along the Woonasquatucket River in RI and is also evident at site 1 (Slack's Tributary, RI) and site 6 (Buckeye Brook, RI). For site 3, the largest source impact based on distance is from a wastewater treatment plant 1 km upstream. No industrial facilities exist upstream of sites 1 and 6. Landfill/waste management facilities are located within 2 km of all three sites but are not hydrologically connected to the sampling locations (Figure S5). Both landfills and wastewater treatment plants are known atmospheric sources of fluorotelomer alcohols and FOSE.<sup>33</sup> Concentrations of N-MeFOSAA, PFUnDA, and PFDoDA were not spatially variable at most sites and only slightly elevated at site 3, consistent with an atmospheric input pathway. We thus infer that this component is most likely attributable to sources from the waste sector.

The third component explains 15% of the variability in PFASs and includes PFPeA, PFOS, and 6:2 FTS. This component is most pronounced at site 2 along the

Woonasquatucket River, within the city of Providence, RI. GIS analysis of the watershed at this site reveals the presence of 14 metal coating/plating industries upstream (Figure 2D, Table S5, and Figure S5). PFOS was historically used as a mist/fume control agent in metal plating, in surface coatings, and as the major component in AFFFs for fighting petroleum-related fires.<sup>25,26,34</sup> Some PFOS applications such as metal plating have been replaced by less stable fluorotelomer-based chemicals such as 6:2 FtS,<sup>35</sup> which will eventually degrade into PFPeA and PFHxA (yields of 1.1 and 1.5% in activated sludge, respectively).<sup>36</sup> It is likely that PFHxA is not included in the cluster because other direct sources can contribute 1 order of magnitude more PFHxA than PFPeA.<sup>37,38</sup> We conclude that the distinct PFAS profile at site 2 can be explained by the metal plating industry.

## ■ IMPLICATIONS

Multivariate statistical tools such as PCA and hierarchical clustering of PFAS profiles combined with data on the hydrological proximity of potential sources are useful for identifying sources of surface water contamination. We find aquatic transport pathways (hydrological distance and river flow directions) are critical for source identification. This contrasts with many other persistent organic pollutants that are primarily transported atmospherically, allowing sources within a radius surrounding the sampling sites to be linked to concentrations.<sup>39</sup> We conclude that the approach demonstrated here for RI and NY/NJ has potential for diagnosing PFAS source contributions in urbanized regions with elevated concentrations and lacking specific information about the magnitude of PFAS discharges from diverse industries. Background PFAS concentrations at most rural sites in this study contain a mix of diverse source signatures that are not statistically distinguishable using these methods. This analysis could be refined in future applications by analyzing additional emerging short-chain PFASs and precursors to develop more unique chemical signatures for specific industries (i.e., those contributing to the first component/cluster).

## ■ ASSOCIATED CONTENT

### S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: [10.1021/acs.estlett.6b00255](https://doi.org/10.1021/acs.estlett.6b00255).

Details on analytical methods, data analysis, and supporting figures and tables ([PDF](#))

## ■ AUTHOR INFORMATION

### Corresponding Author

\*E-mail: [xmzhang@seas.harvard.edu](mailto:xmzhang@seas.harvard.edu). Telephone: 617-495-2893.

### Notes

The authors declare no competing financial interest.

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## ■ REFERENCES

- (1) Grandjean, P.; Andersen, E. W.; Budtz-Jorgensen, E.; Nielsen, F.; Mølbak, K.; Weihe, P.; Heilmann, C. Serum vaccine antibody concentrations in children exposed to perfluorinated compounds. *Jama-J. Am. Med. Assoc.* **2012**, *307*, 391–397.
- (2) Post, G. B.; Louis, J. B.; Lippincott, R. L.; Procopio, N. A. Occurrence of Perfluorinated Compounds in Raw Water from New Jersey Public Drinking Water Systems. *Environ. Sci. Technol.* **2013**, *47*, 13266–13275.
- (3) Castiglioni, S.; Valsecchi, S.; Polesello, S.; Rusconi, M.; Melis, M.; Palmiotti, M.; Manenti, A.; Davoli, E.; Zuccato, E. Sources and fate of perfluorinated compounds in the aqueous environment and in drinking water of a highly urbanized and industrialized area in Italy. *J. Hazard. Mater.* **2015**, *282*, 51–60.
- (4) de Solla, S. R.; De Silva, A. O.; Letcher, R. J. Highly elevated levels of perfluoroctane sulfonate and other perfluorinated acids found in biota and surface water downstream of an international airport, Hamilton, Ontario, Canada. *Environ. Int.* **2012**, *39*, 19–26.
- (5) Hurley, S.; Houtz, E. F.; Goldberg, D.; Wang, M.; Park, J.; Nelson, D. O.; Reynolds, P.; Bernstein, L.; Anton-Culver, H.; Horn-Ross, P.; Petreas, M. Preliminary Associations between the Detection of Perfluoroalkyl Acids (PFAAs) in Drinking Water and Serum Concentrations in a Sample of California Women. *Environ. Sci. Technol. Lett.* **2016**, *3*, 264.
- (6) Happonen, M.; Koivusalo, H.; Malve, O.; Perkola, N.; Juntunen, J.; Huttula, T. Contamination risk of raw drinking water caused by PFOA sources along a river reach in south-western Finland. *Sci. Total Environ.* **2016**, *541*, 74–82.
- (7) Wang, Z. Y.; Cousins, I. T.; Scheringer, M.; Hungerbuhler, K. Fluorinated alternatives to long-chain perfluoroalkyl carboxylic acids (PFCAs), perfluoroalkane sulfonic acids (PFSAs) and their potential precursors. *Environ. Int.* **2013**, *60*, 242–248.
- (8) U.S. Environmental Protection Agency. Facility Registry Service (FRS) (<http://www.epa.gov/enviro/epa-frs-facilities-state-single-file-csv-download>) (accessed November 2015).
- (9) U.S. Environmental Protection Agency. Toxics Release Inventory (TRI) Program (<https://www.epa.gov/toxics-release-inventory-tri-program>) (accessed May 2016).
- (10) Johnson, G. W.; Ehrlich, R.; Full, W.; Ramos, S. *Principal components analysis and receptor models in environmental forensics*; Academic Press: San Diego, 2002.
- (11) Benskin, J. P.; Ahrens, L.; Muir, D. C. G.; Scott, B. F.; Spencer, C.; Rosenberg, B.; Tomy, G.; Kylin, H.; Lohmann, R.; Martin, J. W. Manufacturing Origin of Perfluoroctanoate (PFOA) in Atlantic and Canadian Arctic Seawater. *Environ. Sci. Technol.* **2012**, *46*, 677–685.
- (12) Zhang, X.; Meyer, T.; Muir, D. C.; Teixeira, C.; Wang, X.; Wania, F. Atmospheric deposition of current use pesticides in the Arctic: snow core records from the Devon Island Ice Cap, Nunavut, Canada. *Environ. Sci. Process Impacts* **2013**, *15*, 2304–11.
- (13) Westgate, J. N.; Wania, F. On the construction, comparison, and variability of airsheds for interpreting semivolatile organic compounds in passively sampled air. *Environ. Sci. Technol.* **2011**, *45*, 8850–7.
- (14) Taniyasu, S.; Kannan, K.; So, M. K.; Gulkowska, A.; Sinclair, E.; Okazawa, T.; Yamashita, N. Analysis of fluorotelomer alcohols, fluorotelorner acids, and short- and long-chain perfluorinated acids in water and biota. *J. Chromatogr. A* **2005**, *1093*, 89–97.

- (15) D'eon, J. C.; Crozier, P. W.; Furdui, V. I.; Reiner, E. J.; Libelo, E. L.; Mabury, S. A. Perfluorinated Phosphonic Acids in Canadian Surface Waters and Wastewater Treatment Plant Effluent: Discovery of a New Class of Perfluorinated Acids. *Environ. Toxicol. Chem.* **2009**, *28*, 2101–2107.
- (16) Helsel, D. R. *Nondetects and data analysis: statistics for censored environmental data*; Wiley-Interscience: Hoboken, NJ, 2005; p xv, p 250.
- (17) ESRI Demographics. Population density in the United States ([http://www.esri.com/data/esri\\_data](http://www.esri.com/data/esri_data)), 2012 (accessed May 2016).
- (18) U.S. Geological Survey StreamStats, version 4 (<http://ssdev.cr.usgs.gov/streamstats/>) (accessed May 2016).
- (19) Sakurai, T.; Serizawa, S.; Kobayashi, J.; Kodama, K.; Lee, J. H.; Maki, H.; Zushi, Y.; Sevilla-Nastor, J. B.; Imaizumi, Y.; Suzuki, N.; Horiguchi, T.; Shiraishi, H. Temporal trends for inflow of perfluoroctanesulfonate (PFOS) and perfluoroctanoate (PFOA) to Tokyo Bay, Japan, estimated by a receptor-oriented approach. *Sci. Total Environ.* **2016**, *539*, 277–85.
- (20) Hansen, K. J.; Johnson, H. O.; Eldridge, J. S.; Butenhoff, J. L.; Dick, L. A. Quantitative characterization of trace levels of PFOS and PFOA in the Tennessee River. *Environ. Sci. Technol.* **2002**, *36*, 1681–1685.
- (21) Nakayama, S.; Strynar, M. J.; Helfant, L.; Egeghy, P.; Ye, X. B.; Lindstrom, A. B. Perfluorinated compounds in the Cape Fear Drainage Basin in North Carolina. *Environ. Sci. Technol.* **2007**, *41*, 5271–5276.
- (22) Konwick, B. J.; Tomy, G. T.; Ismail, N.; Peterson, J. T.; Fauver, R. J.; Higginbotham, D.; Fisk, A. T. Concentrations and patterns of perfluoroalkyl acids in Georgia, USA surface waters near and distant to a major use source. *Environ. Toxicol. Chem.* **2008**, *27*, 2011–2018.
- (23) Nakayama, S. F.; Strynar, M. J.; Reiner, J. L.; Delinsky, A. D.; Lindstrom, A. B. Determination of Perfluorinated Compounds in the Upper Mississippi River Basin. *Environ. Sci. Technol.* **2010**, *44*, 4103–4109.
- (24) Lasier, P. J.; Washington, J. W.; Hassan, S. M.; Jenkins, T. M. Perfluorinated chemicals in surface waters and sediments from northwest Georgia, USA, and their bioaccumulation in *Lumbriculus variegatus*. *Environ. Toxicol. Chem.* **2011**, *30*, 2194–201.
- (25) Paul, A. G.; Jones, K. C.; Sweetman, A. J. A first global production, emission, and environmental inventory for perfluoroocane sulfonate. *Environ. Sci. Technol.* **2009**, *43*, 386–392.
- (26) Armitage, J. M.; Schenker, U.; Scheringer, M.; Martin, J. W.; MacLeod, M.; Cousins, I. T. Modeling the Global Fate and Transport of Perfluoroctane Sulfonate (PFOS) and Precursor Compounds in Relation to Temporal Trends in Wildlife Exposure. *Environ. Sci. Technol.* **2009**, *43*, 9274–9280.
- (27) U.S. Census Bureau. Urban And Rural Definitions (<http://www.census.gov/population/censusdata/urdef.txt>), 1995 (accessed June 2016).
- (28) Pistocchi, A.; Loos, R. A Map of European Emissions and Concentrations of PFOS and PFOA. *Environ. Sci. Technol.* **2009**, *43*, 9237–9244.
- (29) Ahrens, L.; Norström, K.; Viktor, T.; Cousins, A. P.; Josefsson, S. Stockholm Arlanda Airport as a source of per- and polyfluoroalkyl substances to water, sediment and fish. *Chemosphere* **2015**, *129*, 33–38.
- (30) Awad, E.; Zhang, X. M.; Bhavsar, S. P.; Petro, S.; Crozier, P. W.; Reiner, E. J.; Fletcher, R.; Tittlemier, S. A.; Braekevelt, E. Long-Term Environmental Fate of Perfluorinated Compounds after Accidental Release at Toronto Airport. *Environ. Sci. Technol.* **2011**, *45*, 8081–8089.
- (31) Filipovic, M.; Woldegiorgis, A.; Norstrom, K.; Bibi, M.; Lindberg, M.; Osteras, A. H. Historical usage of aqueous film forming foam: A case study of the widespread distribution of perfluoroalkyl acids from a military airport to groundwater, lakes, soils and fish. *Chemosphere* **2015**, *129*, 39–45.
- (32) Government of Canada. Chemical Substances: Long-Chain (C9-C20) Perfluorocarboxylic Acids (PFCAs), Their Salts, and Their Precursors (<http://www.chemicalsubstanceschimiques.gc.ca/challenge-defi/summary-sommaire/pfcaps-apfcsp-eng.php>) (accessed November 2015).
- (33) Ahrens, L.; Shoeib, M.; Harner, T.; Lee, S. C.; Guo, R.; Reiner, E. J. Wastewater Treatment Plant and Landfills as Sources of Polyfluoroalkyl Compounds to the Atmosphere. *Environ. Sci. Technol.* **2011**, *45*, 8098–8105.
- (34) Moody, C. A.; Field, J. A. Perfluorinated surfactants and the environmental implications of their use in fire-fighting foams. *Environ. Sci. Technol.* **2000**, *34*, 3864–3870.
- (35) Persistent Organic Pollutants Review Committee of UNEP. Technical paper on the identification and assessment of alternatives to the use of perfluorooctane sulfonic acid, its salts, perfluorooctane sulfonate and their related chemicals in open applications. UNEP/POPS/POPRC.8/INF/17/Rev.1; 2012.
- (36) Wang, N.; Szostek, B.; Buck, R. C.; Folsom, P. W.; Sulecki, L. M.; Gannon, J. T. 8–2 Fluorotelomer alcohol aerobic soil biodegradation: Pathways, metabolites, and metabolite yields. *Chemosphere* **2009**, *75*, 1089–1096.
- (37) Dauchy, X.; Boiteux, V.; Rosin, C.; Munoz, J. F. Relationship Between Industrial Discharges and Contamination of Raw Water Resources by Perfluorinated Compounds: Part II: Case Study of a Fluorotelomer Polymer Manufacturing Plant. *Bull. Environ. Contam. Toxicol.* **2012**, *89*, 531–536.
- (38) Butenhoff, J. L.; Kennedy, G. L., Jr.; Frame, S. R.; O'Connor, J. C.; York, R. G. The reproductive toxicology of ammonium perfluoroctanoate (APFO) in the rat. *Toxicology* **2004**, *196*, 95–116.
- (39) Khairy, M.; Muir, D.; Teixeira, C.; Lohmann, R. Spatial Distribution, Air-Water Fugacity Ratios and Source Apportionment of Polychlorinated Biphenyls in the Lower Great Lakes Basin. *Environ. Sci. Technol.* **2015**, *49*, 13787–97.